

KeyTec® TR-FRET STK discovery kinase kit



CAT.&Size: A1080002S (1,000 tests)

VKEYBIO-01-2025

For Research Use Only

Storage at: 2-8 °C

Not For Diagnostic Or Therapeutic Use

KeyTec® TR-FRET Serine/Threonine Discovery Kinase Kit Technical Manual

1. Introduction

KeyTec® TR-FRET STK discovery kinase kit is designed for measuring the Serine/threonine kinase (STKs) activity. Serine/threonine kinase family comprises multiple members. This assay kit has been validated for activity measurement of over 150 Serine/threonine kinases (For more details, please visit our official website or contact our technical support team). The assay provides three types of biotinylated substrates(S1/S2/S3), Streptavidin-HX¹, a phospho STK-specific antibody labeled with KeyTec® TR-FRET Solar Eu² and detection reagent. These components enable the quantification of kinase activity by detecting the phosphorylated substrates.

This assay is based on a sandwich immunoassay method using KeyTec® TR-FRET technology, offering a simple, rapid, highly specific and sensitive, as well as reproducible detection process. The principle, outlined in Figure 1, involves two main phases:

Kinase Reaction: The specific substrate is phosphorylated by the target kinase in the optimized reaction buffer with ATP.

TR-FRET Detection: Adding an antibody and Streptavidin-HX containing EDTA (to terminate the kinase reaction) that specifically binds to the phosphorylated substrate. This binding brings the TR-FRET donor and acceptor into close proximity, enabling resonance energy transfer (RET) upon excitation and generating a TR-FRET signal. The intensity of TR-FRET signal is directly proportional to the level of phosphorylated substrate.

*1 KeyTec® TR-FRET HX: TR-FRET Acceptor
 *2 KeyTec® TR-FRET Solar Eu: TR-FRET Donor

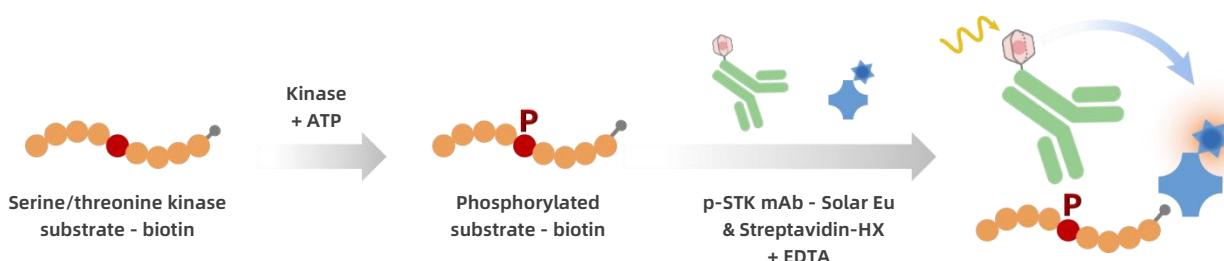


Figure1. The principle of KeyTec® TR-FRET kinase activity detection

2. Components

Components	Storage	A1080002S (1,000 tests ^{*3})
p-STK mAb - Solar Eu (Lyophilized)	2-8 °C	1 vial of 1,000 tests
Streptavidin - HX (Lyophilized)	2-8 °C	2 vials of 840 pmoles A1020024S
STK Substrate 1 - biotin (Lyophilized)	2-8 °C	1 vial of 25 nmoles A1080008S
STK Substrate 2 - biotin (Lyophilized)	2-8 °C	1 vial of 25 nmoles A1080009S
STK Substrate 3 - biotin (Lyophilized)	2-8 °C	1 vial of 25 nmoles A1080010S
DTT (Lyophilized)	2-8 °C	1 vial of 120 µmoles A1010022S
MgCl₂ (1 M)	2-8 °C	1 vial of 1.5 mL A1010024S
CaCl₂ (1 M)	2-8 °C	1 vial of 1.5 mL A1010043S
Kinase Enzymatic buffer (5X)	2-8 °C	1 bottle of 10 mL A1010018S
Kinase Detection buffer (with EDTA)	2-8 °C	1 bottle of 20 mL A1010019S

*3 Tests refer to the number of assay wells that can be performed in 96-well or 384-well plates with 20 μ L in total reaction volume. The reagents of the kit are suggested to use as recommended.

3. Storage

- ◆ Store all reagents according to the recommended conditions. The products are stable for one year from the date of receipt.
- ◆ After thawing, aliquot the stock into single-use volumes (recommended minimum: 10 μ L) to avoid repeated freeze-thaw cycles. Store these aliquots at -60 °C and below.

4. Required Components (Not Supplied)

Material	Brand	Catalog
ATP	Bidepharmatech	BD112724
AMP (5 mM)	Sigma-Aldrich	A1752
cGMP (1 mM)	Sigma-Aldrich	G6129
Lipid activator	Sigma-Aldrich	20-133
Microplate	VKEY-BIO	M2000102N
Top sealing film	VKEY-BIO	M1000102N
Microplate Reader with TR-FRET module	TECAN	Infinite® 200 PRO

5. Procedure

5.1 Reaction system

Phase	Component	Volume ^{*4} (20 µL)
Kinase reaction (10 µL)	Test samples or positive control	2 µL ^{*5}
	Kinase	4 µL ^{*5}
	Serine/threonine kinase substrate - biotin	4 µL (Pre-mix) ^{*5}
	ATP	
TR-FRET detection (10 µL)	p-STK mAb - Solar Eu ^{*6}	5 µL
	Streptavidin - HX ^{*6}	5 µL

^{*4} Recommended Format: Shallow-well 384-well microplate; For 96-well or 1536-well microplates, proportionally scale the reaction system.

^{*5} The proportions of test sample, kinase, substrate and ATP may be adjusted, as long as the concentrations stay the same.

^{*6} It is recommended to premix p-STK mAb - Solar Eu and Streptavidin - HX, add 10 µL pre-mix solution in TR-FRET detection.

5.2 Reagent preparation

- Before use, equilibrate all reagents to RT.
- For lyophilized powder, centrifuge to collect the powder to the bottom (850 xg, 1-2minutes).** It is normal for the fragmentation of lyophilized powder.
- Use the provided/recommended buffers to prepare sample and detection reagents.
- Prepare sample and detection reagent according to the kit technical manual.
- Prepare all reagents immediately before use, unless otherwise specified in the "Working Solution Preparation" section.
- To avoid the degradation of active components, keep 1X Kinase Enzymatic buffer (containing supplemental components) on ice.
- To minimize the aggregation or non-specific binding, It is recommended to add surfactants in Kinase Enzymatic buffer (5X). Notes: Kinase Enzymatic buffer (5X)

contained 0.05% BSA.

- ♦ Avoid vigorous mixing of all reagents.

6. Working Solution Preparation

6.1 Enzyme reaction buffer

- ♦ Prepare 1x enzyme reaction buffer: dilute 1 volume of 5X Kinase Enzymatic buffer and kinase supplements required (e.g., MgCl₂, CaCl₂, DTT etc.) with 4 volume of ultrapure water.
- ♦ **DTT stock solution:** Before reconstitution, centrifuge DTT to pellet the powder to the bottom (850 × g, 1-2 minutes). Add the recommended volume of ultrapure water to the powder and mix gently as below. Aliquot DTT stock solution into single-use tubes to avoid repeated freeze-thaw cycles. These aliquots can be stored at -60°C.

DTT Powder	Buffer	Volume	Stock Concentration
120 µmole/vial	Ultra-pure water	120 µL/vial	1 M
600 µmole/vial	Ultra-pure water	600 µL/vial	1 M

6.2 Test sample or positive control preparation

- ♦ Dilute the test samples to working concentration (5X). For example, if the final test sample concentration is 10 nM in 10 µL of kinase reaction system, dilute test samples to working concentration (50 nM) using 1X enzyme reaction buffer and add 2 µL/well of test samples. The percentage of DMSO must not exceed 2% for serial dilution.

6.3 Substrate and ATP pre-mix solution

- ♦ **ATP stock solution:** Prepare 1 mM ATP stock solution with ultrapure water, aliquot it into single-use tubes to avoid repeated freeze-thaw cycles. These aliquots can be stored at -80°C for up to 3 months.

- ◆ **Substrate stock solution:** Before reconstitution, centrifuge STK substrate - biotin to pellet the powder to the bottom (850×g, 1-2 minutes). Add the recommended volume of ultrapure water to the powder and mix gently as below.

Substrate Powder	Buffer	Volume	Stock Concentration
25 nmoles/vial	Ultra-pure water	500 µL/vial	50 µM

- ◆ **Substrate and ATP pre-mix solution(2.5X):** Prepare 1 mL substrate and ATP pre-mix Solution(2.5X), dilute 50 µL of STK substrate 1 - biotin stock solution and 12.5 µL of ATP stock solution with 937.5 µL of 1x enzyme reaction buffer.

Component	Final Conc. µM	Working Conc. µM	Stock Conc. µM	Dilution	Stock Vol. µL	1X Enzyme Reaction Buffer µL	Total Vol. µL
Substrate - biotin	1.0	2.5	50	20	50	937.5	1,000
ATP	5	12.5	1,000	80	12.5		

6.4 Kinase solution

- ◆ Dilute the kinase to working concentration (2.5X). For example, if the final kinase concentration is 1 ng/µL in 10 µL of kinase reaction system, dilute 100 ng/µL to working concentration (2.5 ng/µL) using 1X enzyme reaction buffer and add 4 µL/well of kinase solution. **Prepare it immediately prior to addition.**

6.5 Detection solution

- ◆ **p-STK mAb - Solar Eu stock solution:** Before reconstitution, centrifuge p-STK mAb - Solar Eu to pellet the powder to the bottom (850×g, 1-2 minutes). Add the recommended volume of Kinase detection buffer to the powder and mix gently as below.

Kit Cat.	Size	Buffer	Volume	Stock Concentration
A1080002S	1,000 tests/vial	Kinase Detection buffer	0.5 mL/vial	10X

- ◆ **p-STK mAb - Solar Eu working solution(1X)**: Dilute 10X p-STK mAb - Solar Eu stock solution to working concentration(1X). For example, prepare 1 mL working solution(1X), dilute 100 μ L of p-STK mAb - Solar Eu stock solution with 900 μ L of kinase detection buffer. Working solution(1X) can be stored at 2-8°C for 24 hours or -60°C for 3 months .
- ◆ **Streptavidin - HX stock solution**: Before reconstitution, centrifuge Streptavidin - HX to pellet the powder to the bottom (850 \times g,1-2 minutes). Add the recommended volume of Ultra-pure water to the powder and mix gently as below.

Component Cat.	Size	Buffer	Volume	Stock Concentration
A1020024S	840 pmoles/vial	Ultra-pure water	42 μ L/vial	20 μ M
A1020024L	8.4 nmoles/vial	Ultra-pure water	420 μ L/vial	20 μ M
A1020024B	40 nmoles/vial	Ultra-pure water	2.0 mL/vial	20 μ M

- ◆ **Streptavidin - HX working solution(1X, 250nM)** : The molar ratio of substrate-biotin: Streptavidin-HX has been optimized at 8:1. Dilute Streptavidin-HX stock solution to working concentration (1X). For example, prepare 0.8 mL working solution(1X) , dilute 10 μ L of Streptavidin - HX stock solution with 790 μ L of kinase detection buffer. Working solution(1X) can be stored at 2-8°C for 24 hours or -60°C for 3 months .

- ◆ **(Optional) p-STK mAb - solar Eu and streptavidin - HX pre-mix solution(0.5X):**
The pre-mix solution should be combined in a 1:1 volume ratio of p-STK mAb-Solar Eu working solution and streptavidin-HX working solution.

Component	Volume	Kinase reaction Conc. (10 μ L in total)	Final Conc. (20 μ L in total)	Working Conc.
Test samples or positive control	2 μ L/well	\	\	\
Kinase	4 μ L/well	0.1 ng/ μ L	\	2.5 ng/ μ L
Substrate - biotin	4 μ L/well (Premix with ATP)	1000 nM	500 nM	2500 nM
Streptavidin - HX (1/8 ⁷)	5 μ L/well	\	62.5 nM ⁷	250 nM ⁷
p-STK mAb - Solar Eu	5 μ L/well	\	\	1X

⁷ Streptavidin - HX / substrate - biotin = 1/8 (Final Conc.) .

7. Procedure

- ◆ Follow the steps in the table below.

		Kinase Group	Negative Control
Kinase reaction	Step 1	2 μ L Test sample (5X) 4 μ L Kinase (2.5X) 4 μ L STK substrate - biotin and ATP pre-mix solution (2.5X)	2 μ L Test sample (5X) 4 μ L Enzymatic buffer 4 μ L STK substrate - biotin and ATP pre-mix solution (2.5X)
	Step 2	Seal the microplate to prevent liquid evaporation, Incubation (RT, 25 °C) for 60 minutes	
TR-FRET detection	Step 3	Remove the top sealing film, add detection solution: 5 μ L p- STK mAb - Solar Eu (1X) 5 μ L Streptavidin - HX (1X) or 10 μ L p-STK mAb - solar Eu and streptavidin - HX pre-mix solution(0.5X)	
	Step 4	Seal the microplate to prevent liquid evaporation, Incubation (RT, 25 °C) for 60 minutes or over-night	
	Step 5	Record the data in microplate reader with top sealing film	

8. Data Analysis

- Calculate the 665 nm/620 nm Ratio (TR-FRET Ratio) and the percentage coefficient of variation (CV %) for each well.

$$\text{TR-FRET Ratio} = \frac{\text{Signal 665 nm}}{\text{Signal 620 nm}} \times 10,000$$

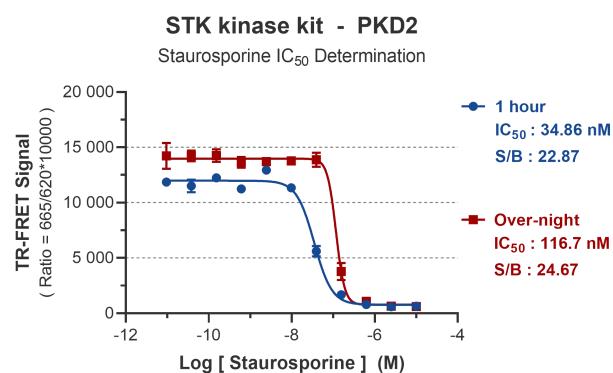
9. Summary

9.1 Results

(1) PKD2

UniProt ID: Q13563

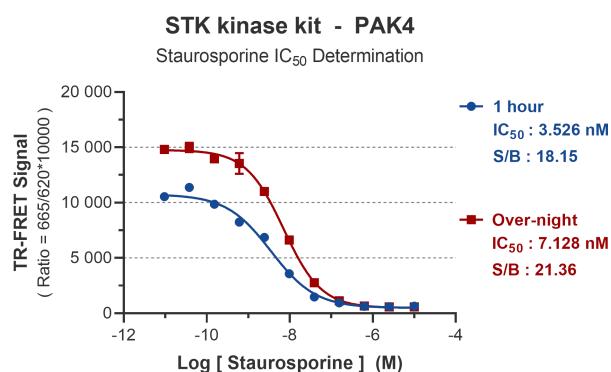
Enzyme concentration	0.015 ng/µL [Final concentration in 10 µL buffer ; 0.15 ng/well]
Incubation time at RT	60 min
Substrate	STK Substrate 1 - biotin, 1 µM [Final concentration in 10 µL buffer]
Streptavidin - HX	62.5 nM [SA/Substrate = 1/8 for final concentration in 20 µL buffer]
Detection antibody	p-STK mAb - Solar Eu, 1X [Working concentration]
ATP concentration	100 µM
Buffer additives	5 mM MgCl ₂ , 1 mM DTT



(2) PAK4

UniProt ID: O96013

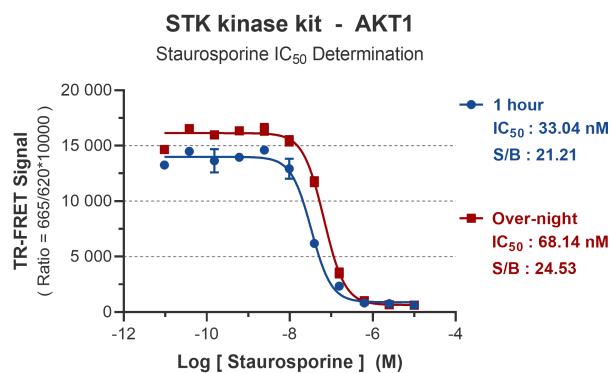
Enzyme concentration	0.1 ng/µL [Final concentration in 10 µL buffer ; 1.0 ng/well]
Incubation time at RT	60 min
Substrate	STK Substrate 2 - biotin, 1 µM [Final concentration in 10 µL buffer]
Streptavidin - HX	62.5 nM [SA/Substrate = 1/8 for final concentration in 20 µL buffer]
Detection antibody	p-STK mAb - Solar Eu, 1X [Working concentration]
ATP concentration	100 µM
Buffer additives	5 mM MgCl ₂ , 1 mM DTT



(3) AKT1

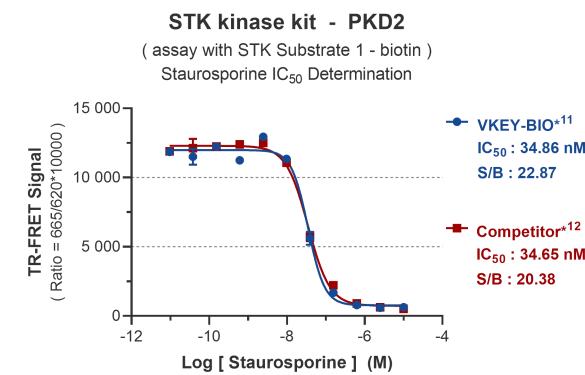
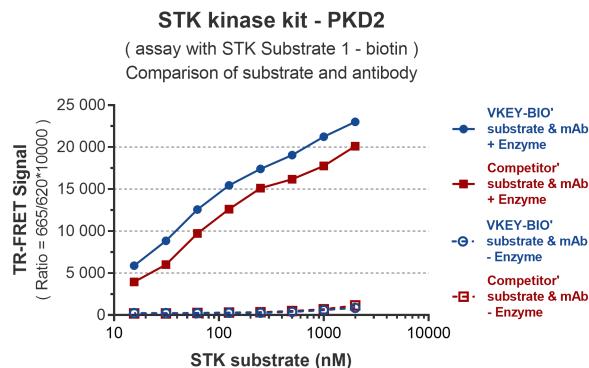
UniProt ID: P31749

Enzyme concentration	0.015 ng/µL [Final concentration in 10 µL buffer ; 0.15 ng/well]
Incubation time at RT	60 min
Substrate	STK Substrate 3 - biotin, 1 µM [Final concentration in 10 µL buffer]
Streptavidin - HX	62.5 nM [SA/Substrate = 1/8 for final concentration in 20 µL buffer]
Detection antibody	p-STK mAb - Solar Eu, 1X [Working concentration]
ATP concentration	100 µM
Buffer additives	5 mM MgCl ₂ , 1 mM DTT

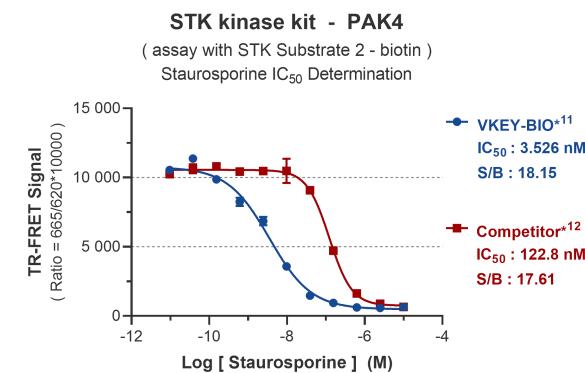
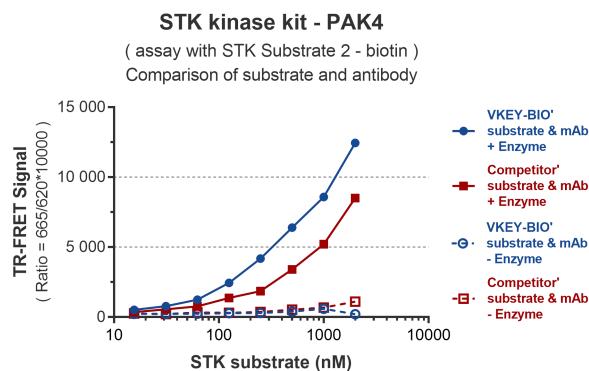


9.2 Performance

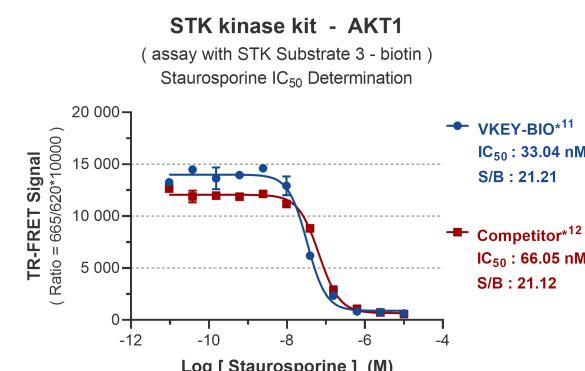
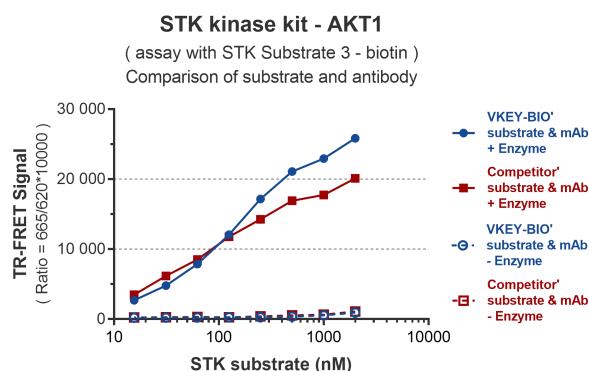
- The comparative data presented below show the performance of the VKEY-BIO KIT and Company C KIT with Substrate 1 under the same experimental conditions.



- The comparative data presented below show the performance of the VKEY-BIO KIT and Company C KIT with Substrate 2 under the same experimental conditions.



- The comparative data presented below show the performance of the VKEY-BIO KIT and Company C KIT with Substrate 2 under the same experimental conditions.



Note: Exemplary data shown. Results are instrument-dependent.

10. Instrument Model and Setting

Vendor	TECAN
Instrument model	Infinite® 200 PRO [Ref. 30050303]
Mode	Fluorescence Top Reading
Excitation filter	320 (25) nm [Ref. 30094454]
Emission filter 1	665 (8.5) nm [Ref. 30094518]
Emission filter 2	620 (10) nm [Ref. 30094505]
Mirror	Dichroic 510
Lag time	150 µs
Integration Time	500 µs
Number of reads	5 or user-defined
Gain	150 or optimal
Z -focus (mm)	Can be calculated on the well giving the highest signal